Guidelines for Euthanasia of Rodents Using Carbon Dioxide

CO2 inhalation is the most common method of euthanasia used at NIH for mice, rats, guinea pigs and hamsters. Although CO2 is generally considered an acceptable euthanasia agent for small animals when properly administered, its acceptability is predicated on a number of critical factors as described in the AVMA Guidelines for the Euthanasia of Animals. The euthanasia method must be appropriate to the research goals, species and age of the animal, approved in the animal study proposal and must conform to the most recent AVMA Guidelines on Euthanasia.

Death by CO2 should be induced as painlessly and quickly as possible. As such, there are a few important aspects of this procedure to consider:

1. Neonatal animals (up to 10 days of age) are resistant to the hypoxia-inducing effects of CO2. Please refer to guidance provided in the ARAC Guidelines for the Euthanasia of Rodent Fetuses and Neonates.

2. Rodents must be euthanized by trained personnel using appropriate technique, equipment and agents.

3. Species should not be mixed during euthanasia.

4. Animals should be euthanized in their home cage whenever possible. Another accepted and common practice is to group animals for euthanasia. Do not overcrowd the chamber. The process of grouping animals immediately prior to euthanasia must provide each individual animal with the ability to make normal postural adjustments.

5. Whenever practical, euthanasia should not be performed in the animal room.

6. The euthanasia chamber should allow animals to be readily visible.

7. Compressed CO2 gas in cylinders is the only recommended source of carbon dioxide as it allows the inflow of gas to the induction chamber to be controlled. Dry ice as a source of CO2 or pre-filled chambers are not acceptable. “Either USP Grade A (medical) or Grade B (industrial) carbon dioxide may be considered acceptable as they each provide a minimum purity for carbon dioxide of 99.0%.”

8. Without pre-charging the chamber, place the animal(s) in the chamber and introduce 100% carbon dioxide. A fill rate of 30% of the chamber volume per minute with carbon dioxide, added to the existing air in the chamber is appropriate to achieve a balanced gas mixture to fulfill the objective of rapid unconsciousness with minimal distress to the animals.

   a. Example: for a 10-liter volume chamber, use a flow rate of 3 liter(s) per minute.
   b. Use the formula in the table below to calculate the appropriate flow rate for your chamber/size.
9. Expected time to unconsciousness is usually within 2 to 3 minutes. Observe each rodent for lack of respiration and faded eye color. Maintain CO₂ flow for a minimum of 1 minute after respiration ceases. If both signs are observed, then remove the rodents from the cage; otherwise continue exposing them to CO₂. If unconsciousness has not yet occurred within 2 to 3 minutes, the chamber fill rate should be checked. The system should also be examined for a defective flow meter, absence of CO₂ supply, and/or leaks. Appropriate CO₂ concentrations and exposure times will prevent unintended recovery.

10. Upon completion of the procedure, death must be confirmed by an appropriate method, such as ascertaining cardiac and respiratory arrest or noting an animal's fixed and dilated pupils. It is important to verify death after CO₂ exposure. If an animal is not dead, CO₂ narcosis must be followed by a secondary method of euthanasia, such as decapitation, bilateral pneumothorax, or cervical dislocation.

11. If a home cage cannot be used, the CO₂ euthanasia chamber should be cleaned between each use and at the end of the day to remove debris or pheromones expressed during the previous euthanasia session. Alternatively, a new/unused container should be used with each group.
References

3. AVMA. may change guidance for CO2 euthanasia in rodents. JAVMA, 2019, 254 (1) 31.

Useful Review

- Biovin GP, Bottomley MA, Dudley ES, Schimi PA, Wyatt CN, Grobe N. Physiological, behavioral, and histological responses of male C57BL/6N mice to different CO2 chamber replacement rates. JAALAS, 2016, 55(4), 451-61.
- Klaunberg BA, O’Malley J, Clark T, Davis JA. Euthanasia of Mouse Fetuses and Neonates. Contemporary Top Lab Anim Sc 2004, 43:(5)29-34.
- Wong D, Makowska IJ, Weary DM. Rat aversion to isoflurane versus carbon dioxide. Biology letters, 2013, 9 (1).

Approved - 09/12/01
Revised - 10/09/02, 10/13/04, 10/10/07, 05/12/10, 05/08/13, 1/25/17, 10/23/2019
Calculating Rodent Euthanasia Chamber Size & Flow Rate

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<tbody>
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<td>Chamber Height</td>
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<td>Chamber Width</td>
<td>5</td>
</tr>
<tr>
<td>Chamber Length</td>
<td>4</td>
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CO₂ Flow Rate Calculations (L/min) will appear below in blue boxes.

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<tbody>
<tr>
<td>Chamber Volume (Cubic Inches)</td>
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<tr>
<td>Chamber Volume (Liters)</td>
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<tr>
<td>10% Displacement Rate (no longer recommended)*</td>
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<td>20% Displacement Rate (no longer recommended)*</td>
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<tr>
<td>30% Displacement Rate (required)*</td>
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*See NIH ARAC Guidelines for Euthanasia of Rodents Using Carbon Dioxide for additional details.