Guidelines for Pain and Distress in Laboratory Animals: Responsibilities, Recognition, and Intervention

Introduction
Animals can experience pain and distress. It is the ethical and legal obligation of all personnel involved with the use of animals in research to reduce or eliminate pain and distress in research animals whenever such actions do not interfere with the research objectives. The Institute/Center Animal Care and Use Committee (IC ACUC) has the delegated responsibility and accountability for ensuring that all animals under their oversight are used humanely and in accordance with a number of Federal Regulations and policies.2,21,29,30,32 Key to fulfilling the responsibilities for both the Principal Investigator (PI) and the IC ACUC are to:

- understand the legal requirements,
- be able to distinguish pain and distress in animals from their normal state,
- relieve or minimize the pain and distress appropriately; and
- establish humane endpoints.

Regulatory Requirements and IC ACUC Responsibilities
The IC ACUC must ensure that all aspects of the animal study proposal (ASP) that may cause more than transient pain and/or distress are addressed; alternatives6 to painful or distressful procedures are considered; that methods, anesthetics, and analgesics to minimize or eliminate pain and distress are included when these methods do not interfere with the research objectives; and that humane endpoints have been established for all situations where more than transient pain and distress can not be avoided or eliminated. Whenever possible, death or severe pain and distress should be avoided as endpoints. A written scientific justification is required to be included in the ASP for any more than transient painful or distressful procedure that cannot be relieved or minimized.

The following items should be considered when developing and evaluating justifications for studies involving increased levels of pain and distress:

- Description of experimental procedure (including conditioning/training plan, when applicable);
- Assurance that less painful/distressful procedures were considered, and, if available, why they were not used;
- Description of monitoring plan for during and after the experiment (including monitoring parameters, monitoring schedule, and assignment of monitoring duties);
- Description of exclusion criteria;
- Description of anticipated clinical effects;
- Description of interventions to minimize pain/distress, including both pharmacological and non-pharmacological methods;
- Description of clear humane endpoints; and
- Assignment of endpoint decision-making responsibilities.

The obligation to reduce pain and distress does not end with the review of the ASP. It is the responsibility of the animal care staff, the research staff, veterinarians and the IC ACUC, to continue to monitor animals for pain, distress, illness, morbidity or mortality during the course of the research study.

If unexpected pain or distress occurs, and is more than an isolated incident, it is the PI’s responsibility to
submit an amendment delineating the unexpected problem and the proposed resolution (e.g., administration of analgesics, lowering the dose of a drug that was administered, etc.). Alternatively, the PI could justify the need for unrelieved pain or distress in the amendment, and in the case of regulated species, in a Column E Justification form.

**Recognition of Pain and Distress**

Animals must be monitored by trained individuals for pain and distress as appropriate for the species, condition, and procedure. Critical to the assessment of the presence or absence of pain or distress is having the ability to distinguish between normal and abnormal animal behavior. This is especially true when dealing with species that often exhibit pain and distress with only subtle changes in their behavior (see Table 1). Therefore, it is critical that the individuals assessing an animal be trained in the species-specific signs of pain and distress, as well as be knowledgeable of the potential outcomes of the procedure, surgery or treatments administered to the animal. Pain and distress scoring is a method to convert subjective animal observations into an objective scoring system which some have found to be helpful in assessing animal behavior.

Whenever more than transient pain or distress is anticipated, preemptive measures should be taken to minimize or prevent the development of pain and/or distress. Following the implementation of preemptive or palliative measures, animals must be monitored to ensure the efficacy of the measures taken and determine if or when additional treatment will be necessary. The extent and frequency of monitoring will depend on the level of post-surgical/procedural pain and/or distress anticipated and the chosen intervention strategy(s). For example, animals undergoing a procedure known to produce no more than minimal/transient pain or distress may be adequately monitored by the daily observation of a trained animal caretaker. Whereas the monitoring of an animal undergoing a procedure known to result in severe pain and/or distress may require more frequent monitoring by a team of trained individuals (e.g., trained animal care staff, technicians, veterinarians, investigators, etc.). Animals undergoing pilot studies or procedures new to the investigator or facility may also require a higher frequency of monitoring and a team approach.

It is ultimately the responsibility of the PI and the personnel conducting the procedure to ensure the timely and adequate identification, monitoring, and documentation of the animals undergoing potentially painful or distressful procedures. Investigators may request the assistance of institute and facility veterinary and technical personnel when monitoring their animals, but all individual(s) responsible for monitoring an animal must be identified prior to conducting the procedure and their accountability clearly delineated and accepted.

Animals should be observed a minimum of once daily or more often based on professional judgment and the research being conducted. The animals should be monitored for expected and unexpected signs of more than transient pain or distress and, if observed, appropriate intervention strategies implemented (e.g., non-pharmacological approaches, analgesics, anesthetics, euthanasia, etc.), unless the withholding of treatment is scientifically justified. Observations and actions taken to relieve pain and/or distress must be documented.

The documentation of monitoring of the animal is important and required. The nature and the frequency of the documentation are dependent on the species and the potential for pain and/or distress. For example, the identification of cages containing animals where a potentially painful or distressful procedure has been performed with a “special observation” cage card has proven helpful in drawing special attention to the animal during the caretaker’s daily health check. Cages containing animals requiring more intensive monitoring should also be appropriately identified and their
monitoring and/or treatments documented either at the room, cage or animal level (e.g., room log, cage card, medical record, etc.), in addition to the investigator’s notations in their laboratory notebook. Documentation must be available to all personnel monitoring the cage or animal (i.e., IC ACUC, veterinarians, animal care staff, etc.).

**Intervention Strategies**

Strategies for the management of pain and distress may include non-pharmacological considerations (e.g., modified housing and husbandry practices, dietary modifications, surgical approaches, desensitization, and acclimation strategies, etc.), pharmacological interventions, or euthanasia. The chosen strategy will vary with the species, the procedure(s) being performed, duration of action needed, route of administration preferred, degree and type of analgesia required, and research being conducted (see Table 2). It is strongly suggested that PIs consult their IC veterinarian during the development of an ASP, prior to its submission to the IC ACUC. This approach has been demonstrated to expedite the ASP approval process.

Excellent resources and formularies are available which provide extensive information on the recognition and alleviation of pain and distress in laboratory animals (See Plumb Veterinary Drug Handbook and other references, below). These resources, coupled with trained and skilled animal care personnel, and the professional judgment of IC and animal facility veterinarians, provide each investigator and their IC ACUC with powerful tools for the recognition and alleviation of pain and distress in laboratory animals.

Preemptive measures should be taken to minimize or prevent the development of pain and/or distress. For example, a skilled surgeon can often minimize tissue trauma which in turn minimizes post-operative pain and distress. The use of ketamine or opioids preemptively, even in low doses has been demonstrated to prevent the development of some forms of pain. In addition, the use of a single dose of a non-steroidal anti-inflammatory agent (NSAID), sustained-release formulations, or other analgesic agent can have a positive effect on the speed with which animals return to normal behavior. It has been repeatedly demonstrated in humans that the provision of effective analgesia reduces the time taken for post-operative recovery.

There are also many pharmacological intervention strategies for the management of pain and distress. Traditional analgesics include local or regional anesthetics, opioids and NSAIDs. Using two or more of classes of these analgesics together or combining these analgesics with nontraditional analgesics such as N-Methyl-D-aspartate (NMDA) antagonists, alpha2-agonists, tramadol, and even the antiepileptic drug gabapentin have been shown in both human and veterinary patients to enhance analgesia and allow a reduction in the use of more powerful analgesics. This approach is called multimodal analgesia. It has the advantage of providing even analgesic dosing thus promoting the animal’s well being.

For procedures in which the pain intensity is anticipated to be high, techniques such as constant rate infusions of local anesthetics and or opioids either systemically, locally at the surgical site or via an epidural catheter and transdermal preparations of drugs provide uninterrupted analgesia and are being used successfully in larger laboratory animals. The analgesic regimen chosen should always be made in consultation with the veterinarian.

**Summary**

The relief of pain and distress in research animals is ethically sound, humane, and promotes good science. The establishment of clear lines of responsibility coupled with appropriate endpoints,
monitoring and intervention strategies are key to the prevention, minimization and/or alleviation of pain and distress in laboratory animals. Several excellent references and formularies are available to the researcher, veterinarian, and husbandry personnel to facilitate their ability to recognize and modulate pain and distress in laboratory animals. Experience has demonstrated that a dynamic, interactive team approach to the recognition and alleviation of pain and distress in laboratory animals yields results that protect animal welfare while promoting good science.

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Revised - 07/14/04; 05/16/07; 07/14/10; 11/14/12, 06/10/2015, 01/23/2019, 12/09/2020

References


### TABLE 1: POTENTIAL SIGNS ASSOCIATED WITH PAIN OR DISTRESS IN RATS, MICE AND RABBITS

<table>
<thead>
<tr>
<th>Potential Signs</th>
<th>Mice</th>
<th>Rats</th>
<th>Rabbits</th>
</tr>
</thead>
<tbody>
<tr>
<td>Decreased Food and Water Consumption</td>
<td>X</td>
<td>X</td>
<td>X</td>
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<tr>
<td>Weight loss</td>
<td>X</td>
<td>X</td>
<td>X</td>
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<tr>
<td>Self-imposed isolation/hiding</td>
<td>X</td>
<td>X</td>
<td>X</td>
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<tr>
<td>Self-mutilation, gnawing at limbs</td>
<td>X</td>
<td>X</td>
<td>X</td>
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<tr>
<td>Rapid Breathing</td>
<td>X</td>
<td>X</td>
<td>X</td>
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<tr>
<td>Opened-Mouth Breathing</td>
<td>X</td>
<td>X</td>
<td>X</td>
</tr>
<tr>
<td>Abdominal Breathing</td>
<td>X</td>
<td>X</td>
<td>X</td>
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<tr>
<td>Grinding Teeth</td>
<td>X</td>
<td>X</td>
<td></td>
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<tr>
<td>Biting/Growling/Aggression</td>
<td>X</td>
<td>X</td>
<td></td>
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<tr>
<td>Increased/Decreased Movement</td>
<td>X</td>
<td>X</td>
<td>X</td>
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<tr>
<td>Unkempt Appearance (Erected, Matted, or Dull Haircoat)</td>
<td>X</td>
<td>X</td>
<td>X</td>
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<tr>
<td>Abnormal Posture/Positioning (e.g., Head-pressing, Hunched Back)</td>
<td>X</td>
<td>X</td>
<td>X</td>
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<tr>
<td>Restless Sleep</td>
<td></td>
<td>X</td>
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<tr>
<td>Tearing (including Porphyria), Lack of Blinking Reflex</td>
<td>X</td>
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<tr>
<td>Dilated Pupils</td>
<td>X</td>
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<tr>
<td>Muscle Rigidity, Lack of Muscle Tone</td>
<td>X</td>
<td>X</td>
<td>X</td>
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<tr>
<td>Dehydration/Skin Tenting/Sunken Eyes</td>
<td>X</td>
<td>X</td>
<td>X</td>
</tr>
<tr>
<td>Twitching, trembling, tremor</td>
<td>X</td>
<td>X</td>
<td>X</td>
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<tr>
<td>Vocalization (Rare)</td>
<td>X</td>
<td>X</td>
<td>X</td>
</tr>
<tr>
<td>Redness or Swelling Around Surgical Site</td>
<td>X</td>
<td>X</td>
<td>X</td>
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<tr>
<td>Increased Salivation</td>
<td>X</td>
<td>X</td>
<td>X</td>
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### TABLE 2: POST PROCEDURAL PAIN POTENTIAL a,b

<table>
<thead>
<tr>
<th>Minimal to Mild Pain c</th>
<th>Mild to Moderate Pain d</th>
<th>Moderate to Severe Pain e</th>
</tr>
</thead>
<tbody>
<tr>
<td>Catheter implantation</td>
<td>Minor laparotomy incisions</td>
<td>Major laparotomy/organ incision</td>
</tr>
<tr>
<td>Tail clipping</td>
<td>Thyroidectomy</td>
<td>Thoracotomy</td>
</tr>
<tr>
<td>Ear notching</td>
<td>Orchidectomy</td>
<td>Heterotopic organ transplantation</td>
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<tr>
<td>Subcutaneous transponder placement</td>
<td>C-section</td>
<td>Vertebral procedures</td>
</tr>
<tr>
<td>Superficial tumor implantation</td>
<td>Hypophysectomy</td>
<td>Burn procedures</td>
</tr>
<tr>
<td>Orbital sinus venotomy</td>
<td>Thymectomy</td>
<td>Trauma models</td>
</tr>
<tr>
<td>Rodent embryo transfer</td>
<td>Embryo transfer in non-rodents</td>
<td>Orthopedic procedures</td>
</tr>
<tr>
<td>Multiple injections</td>
<td>Bone marrow collection</td>
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<tr>
<td>Non-corneal ocular procedures</td>
<td>Corneal procedures</td>
<td></td>
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<tr>
<td>Intracerebral electrode implantation</td>
<td></td>
<td></td>
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<tr>
<td>Vasectomy</td>
<td></td>
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<tr>
<td>Vascular access port implantation</td>
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<td></td>
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<tr>
<td>Craniotomy (periosteal pain)</td>
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<td></td>
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<tr>
<td>Superficial lymphadenectomy</td>
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</tbody>
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**a** Table adapted from “Guidelines for the Assessment and Management of Pain in Rodents and Rabbits”. 2006, American College of Laboratory Animal Medicine

**b** The analgesia and monitoring required may vary due to a number of factors; such as the invasiveness of the procedure, degree of tissue trauma, surgical time, skill of the surgeon, and the tissues or organs involved.

**c** Post procedural pain relief for minimal to mild pain may be adequately addressed with preemptive analgesia, tissue infiltration with a long acting local anesthetic, and a single dose of a long acting NSAID or mixed opioid agonist-antagonist, or other agent.

**d** Post procedural pain relief for mild to moderate pain may be adequately addressed with tissue infiltration with a long-acting local anesthetic combined with one or more doses of a long acting NSAID and/or an opioid or other agent in addition to pre-emptive analgesic administration.

**e** Post procedural pain relief for moderate to severe pain should encompass multimodal analgesia (e.g., combining a pure opioid agonist with a NSAID, tissue infiltration with a long-acting local anesthetic, etc.)